

Evidence of an α_2 -macroglobulin-like molecule in plasma of *Salamandra salamandra*

Structural and functional similarity with human α_2 -macroglobulin

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A high- M_r (M_r 750 000) α_1 -macroglobulin, obtained from *Salamandra salamandra*, is described. Salamander α_1 -macroglobulin is composed of two monomers of equal M_r , which are composed of two polypeptide chains, each of M_r 180 000, linked by disulfide bonds. The molecular parameters of this protein, its binding to trypsin and inactivation by methylamine suggest that salamander α_1 -macroglobulin is closely related to human α_2 -macroglobulin and to other related proteins described in the animal kingdom.

α_1 -Macroglobulin; α_2 -Macroglobulin; Trypsin protecting activity; Antiprotease; (Salamander, Urodele)

1. INTRODUCTION

α_2 -Macroglobulin (α_2M) is a major protease inhibitor of human serum [1]. This protein has an M_r of 720 000 and is composed of two monomers of M_r 360 000, non-covalently linked [2]. Four thiol ester bonds have been described, which can be cleaved by small amines, like methylamine [3–6]. α_2M inhibits the four classes of endoproteases by a unique mechanism of steric hindrance, not involving the active site [7–9].

Analogues of human α_2M have been found in the sera and plasma of other mammalian species, i.e. in rats, rabbits and dogs [10–13]. In those species,

α_1 - and α_2 -macroglobulins can be observed depending on the physiological conditions (normal or acute-phase) [14–16]. Macroglobulins have also been described for the bird [17,18], reptile [17], frog [17–19] and fish [20,21] as well as for the horseshoe crab *Limulus limulus* [17,22,23]. The molecular properties of a urodele salamander antiprotease (α_1M) are described here [24].

2. MATERIALS AND METHODS

Salamander antiprotease α_1M (S- α_1M) was isolated from plasma of 20 male salamanders as in [24], its native M_r of 750 000 being determined as in [25], and stored at -80°C in 0.01 M phosphate buffer 0.15 M NaCl, pH 7.2 (buffer A). Human α_2M (H- α_2M) was prepared from plasma of seven donors according to Barrett et al. [2], except that the last stage of immuno-adsorption was omitted. Storage conditions were as for S- α_1M . Porcine trypsin (Sigma) was dissolved in 1 mM HCl to a concentration of 6.6×10^{-7} M and was 60% active [26].

Protein concentration was determined by the

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Abbreviations: α_2M , α_2 -macroglobulin; α_1M , α_1 -macroglobulin; STI, soybean trypsin inhibitor; BSA, bovine serum albumin; TAME, *N*- α -p-toluenesulfonyl-L-arginine methyl ester; 2ME, 2-mercaptoethanol; PAGE, polyacrylamide gel electrophoresis

technique of Lowry et al. [27], using BSA as reference. Esterase activity of α M-trypsin complexes was studied in buffer B consisting of 46 mM Tris-HCl, 11 mM CaCl_2 , pH 8.1, with 10 mM TAME as substrate. Buffer C consisted of 20 mM Tris-HCl, pH 8.0. STI (Sigma) was 6.6×10^{-6} M in buffer A. Stock solution of methylamine (Sigma) was 200 mM in buffer C. Electrophoresis was performed as described [24], except that samples to be denatured were incubated for 45 min at 37°C to prevent autolysis of α 2M or α 1M [2].

3. RESULTS

The M_r values of S- α 1M and H- α 2M subunits were determined by PAGE (fig.1). The results demonstrate that the monomers of both molecules behave similarly, i.e. giving one band of M_r 360 000. Under reducing conditions, the polypeptide chains of S- α 1M and H- α 2M had nearly the same M_r (M_r 180 000 for S- α 1M and 170 000 for H- α 2M). These data are in good agreement with

those given in the literature [2]: the half-molecule of H- α 2M (M_r 360 000) is composed of two polypeptide chains (M_r 180 000). The capacity of S- α 1M and H- α 2M to complex trypsin without affecting its esterolytic activity was investigated using the technique of Ganrot [9] (fig.2). In the absence of methylamine, both molecules exhibited the same binding to trypsin, as demonstrated by their protection of trypsin from inhibition by STI. In contrast, such protection was not found after treatment with methylamine, thus indicating that the amine prevented the formation of the complexes between the macroglobulins and trypsin, presumably by hydrolysing the thiol ester (fig.2). Thus, S- α 1M appears, both structurally and functionally, as a homologue of H- α 2M.

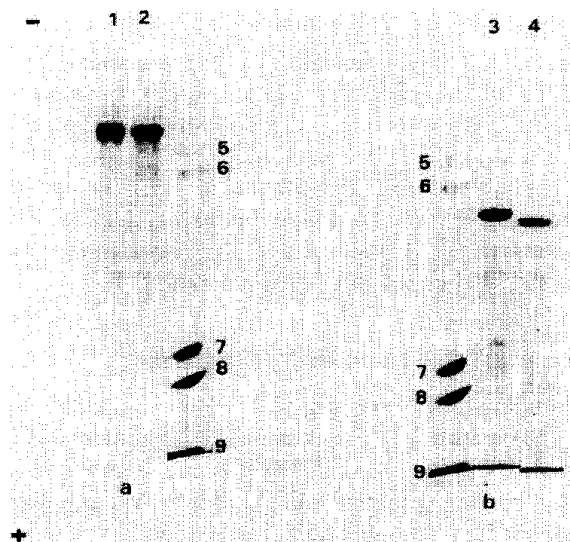


Fig.1. Polyacrylamide (7%) gel electrophoresis of salamander α 1M and human α 2M, in the presence of 2% SDS (a) and 2% SDS + 5% 2ME (b). Stacking gel: 5% in acrylamide. Tracks 1,3: salamander α 1M (30 μg); tracks 2,4: human α 2M (30 μg). Voltage, 150 V. Time of migration, 5 h. Kit migrating position: (5) thyroglobulin (330 kDa); (6) ferritin (220 kDa); (7) albumin (67 kDa); (8) catalase (60 kDa); (9) lactate dehydrogenase (36 kDa).

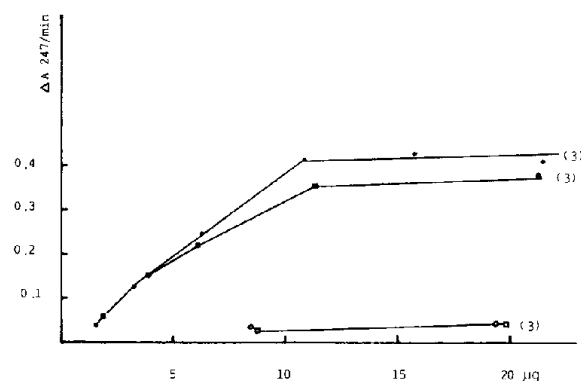


Fig.2. Esterolytic activity of S- α 1M or H- α 2M in the presence or absence of 0.2 M methylamine, followed by addition of STI. For experiments performed in the absence of methylamine, 0.45 μg active trypsin was mixed with increasing amounts (from 0 to 30 μg) of α 1M (●) or α 2M (■). After incubation for 15 min at 37°C , 1.5 μg STI (10 μl) was added. After a further incubation for 10 min at 37°C , this mixture was added to a cuvette containing 1300 μl buffer B and 150 μl TAME (10 mM in distilled water). The increase in absorbance at 247 nm was recorded for 4 min at 25°C . For experiments performed in the presence of methylamine, 8.1 and 19.3 μg α 1M (○) or 8.5 and 19.9 μg α 2M (□), diluted in buffer C, were incubated for 180 min at room temperature with 100 μl of 200 mM methylamine in buffer C. The mixture was then added to 0.45 μg trypsin as described before. The rest of the manipulation was as described above. The points on each line represent the mean of 3 experiments.

4. DISCUSSION

α -Macroglobulins have been described in all the classes of vertebrates [17] and even in the invertebrate *L. limulus* [17,22,23]. Feldman and Pizzo [19] have shown that frog α 1M has the same M_r (M_r 730000) as human α 2M and that it is composed of four polypeptide chains of M_r 180000, associated in disulfide-bonded pairs. Salamander is a Urodele amphibian, more primitive than frog on the phylogenetic scale, and one of the earliest tetrapods to demonstrate viviparity [24]. Our results allow one to suggest that S- α 1M has the same molecular characteristics as those in anurans such as frog [19], and different from those in fish [17]. S- α 1M is composed of four polypeptide chains of M_r 180000, as described in vertebrate classes, except in fish where α Mh is composed of two chains of M_r 105000 and 90000 [20].

The reaction of trypsin with S- α 1M and H- α 2M could not be inhibited by STI, while a major activity against TAME was maintained. The stoichiometric ratio between trypsin and S- α 1M was similar to that given by Feldman and Pizzo [19] for frog. In contrast, trypsin treated with methylamine-S- α 1M complex remained highly sensitive to STI. As expected [28], the same behaviour was observed with the human control α 2M. Thus, these data are consistent with the hypothesis that *Salamandra salamandra* α 1M, a protein of M_r 750000 is the homologue of human α 2M and of similar proteins in other animal species.

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REFERENCES

- [1] Starkey, P.M. and Barrett, A.J. (1977) in: Proteinases in Mammalian Cells and Tissues, pp.663-699, Elsevier, Amsterdam, New York.
- [2] Barrett, A.J., Brown, A. and Sayers, C.A. (1979) Biochem. J. 181, 401-418.
- [3] Steinbuch, M., Pejaudier, L., Quentin, M. and Martin, V. (1968) Biochim. Biophys. Acta 154, 228-231.
- [4] Swenson, R.P. and Howard, J.B. (1971) Proc. Natl. Acad. Sci. USA 66, 4313-4316.
- [5] Swenson, R.P. and Howard, J.B. (1980) J. Biol. Chem. 255, 8087-8091.
- [6] Howard, J.B. (1981) Proc. Natl. Acad. Sci. USA 78, 2235-2239.
- [7] Haverback, B.J., Dye, B., Bundy, H.F., Wirtshafter, S.K. and Edmonson, H.A. (1962) J. Clin. Invest. 41, 972-980.
- [8] Barrett, A.J., Starkey, P.M. and Munn, E.A. (1974) in: Bayer Symposium V, Proteinase Inhibitors, pp.72-77, Springer, New York.
- [9] Ganrot, P.O. (1966) Clin. Chim. Acta 14, 493-501.
- [10] James, K. (1965) Immunology 8, 55-61.
- [11] Bjork, I., Lindblom, T. and Lindahl, P. (1985) Biochemistry 24, 2653-2660.
- [12] Dangott, J. and Cunningham, L.W. (1982) Biochem. Biophys. Res. Commun. 107, 1243-1251.
- [13] Ganrot, K. (1973) Biochim. Biophys. Acta 322, 62-67.
- [14] Gordon, A.H. (1976) Biochem. J. 159, 643-650.
- [15] Ganrot, K. (1973) Biochim. Biophys. Acta 295, 245-251.
- [16] Heim, W.G. (1968) Nature 217, 1057-1059.
- [17] Starkey, P.M. and Barrett, A.J. (1982) Biochem. J. 205, 91-95.
- [18] Nagase, H., Harris, E.D., Woeddnner, J.F. and Brew, K. (1983) J. Biol. Chem. 258, 7481-7489.
- [19] Feldman, S.R. and Pizzo, S.V. (1985) Biochemistry 24, 2569-2575.
- [20] Starkey, P.M. and Barrett, A.J. (1982) Biochem. J. 205, 105-115.
- [21] Starkey, P.M., Fletcher, T.C. and Barrett, A.J. (1982) Biochem. J. 205, 97-104.
- [22] Quigley, J.P. and Armstrong, P.B. (1983) J. Biol. Chem. 258, 7903-7906.
- [23] Quigley, J.P. and Armstrong, P.B. (1985) J. Biol. Chem. 260, 12715-12719.
- [24] Badet, M.T., Chateaufreynaud-Duprat, P., Mouches, C., Bové, J.M. and Metivaud, J. (1980) Ann. Inst. Pasteur/Immunol. 131D, 57-62.
- [25] Sallenave, J.M. (1986) Thèse 3e cycle Biologie et Physiologie Animales, Bordeaux I.
- [26] Chase, T. and Shaw, E. (1967) Biochem. Biophys. Res. Commun. 29, 508.
- [27] Lowry, O.H., Rosebrough, N.J., Farr, A.L. and Randall, R.J. (1951) J. Biol. Chem. 193, 265-275.
- [28] Gonias, S.L., Reynolds, J.A. and Pizzo, S.V. (1982) Biochim. Biophys. Acta 705, 306-314.